Acta Zoologica Academiae Scientiarum Hungaricae 55 (2), pp. 187–197, 2009

GENETIC DIVERSITY IN PERIPHERAL AND CENTRAL POPULATIONS OF RUSTY-NECKLACED PARTRIDGE (ALECTORIS MAGNA) BASED ON MITOCHONDRIAL AND MICROSATELLITE DNA

HUANG, Z. H.¹, LIU, N. F.^{2*}, CHEN, Y. K.² and XIAO, Y. A.¹

¹School of Life Sciences, Jinggangshan University, Ji'an, Jiangxi, 343009, China E-mail: hzhow@163.com ²School of Life Sciences, Lanzhou University, Lanzhou, Gansu, 730000, China

Though it has been long presumed that peripheral populations tend to exhibit low levels of genetic diversity due to isolation and genetic drift, results of empirical investigation remain ambiguous. Some rusty-necklaced partridge (*Alectoris magna*) populations have expanded their present ranges, resulting in several peripheral populations, due to recent deforestation by human beings in Northwestern China. On the basis of mitochondrial DNA control-region and microsatellite DNA data, we compare the genetic diversity (π -, H-, H_0 -, and H_E -values) between three peripheral populations and five central populations. Maternal and biparental DNA markers indicated accordantly genetic diversity. Compared to central populations, the peripheral populations exhibited lower genetic diversity. The low genetic variability of the three peripheral populations appeared to result partly from isolation and natural selection.

Key words: *Alectoris magna*, peripheral population, genetic diversity, mitochondrial DNA, microsatellite

INTRODUCTION

Geographically peripheral populations are more likely to be imperiled than central populations. They tend to occur in less suitable environments and are often isolated from more central and continuous populations (LESICA & ALLENDORF 1995). Many theoretical works have revealed that genetic mechanisms such as inbreeding or genetic drift in small population caused by genetic bottlenecks and founder effects are important factors in reducing genetic variability (BARRETT & KOHN 1991). Genetic diversity is expected to be lower in peripheral populations than in central populations (CASSEL & TAMMARU 2003, ECKERT *et al.* 2008), due to genetic drift (NEI *et al.* 1975, HARTL & CARK 1997). Central populations are usually large, continuous and occupy favorable habitats. Peripheral populations, by contrast, can be more or less isolated, fragmented, and be subject to a more variable physical environment (LESICA & ALLENDORF 1995). Accordingly, peripheral populations will often experience different selection pressures than central popula-

* The first two authors contributed equally to this work.

tions, which may lead to lower genetic divergence (KIRPATRICK & BARTON 1997). Genetic differences are most likely to occur in populations that become isolated at the periphery of the range (LESICA & ALLENDORF 1995, SAFRIEL *et al.* 1994). Empirical evidence supporting this hypothesis remains ambiguous (GARNER *et al.* 2004, HUANG *et al.* 2005). Some data support the hypothesis that peripheral populations exhibit lower genetic diversity (LAMMI *et al.* 1999, HOU *et al.* 2002, WANG *et al.* 2001), while others show no such relationship (TIGERSTEDT 1973, WENDEL & PARKS 1985, PETITET *et al.* 1998). There is thus practical need for descriptive studies.

Rusty-necklaced partridge (*Alectoris magna*, Gallifromes, Phasianidae) is found in Qinghai, Ningxia and Gansu provinces, patchily distributed in dry and open rocky mountains (LIU 1992), with two described subspecies: *A. magna magna* (the Chaidamu Basin) and *A. magna lanzhouensis* (the Lanzhou Basin and the Liupanshan Mountain) (LIU *et al.* 2004). The partridge is representative species of arid and semiarid environments in northwestern China (HUANG *et al.* 2007*a*). Forest and farmland are generally avoided. Most forest has disappeared in the Gansu Province because of deforestation and cultivation by human beings. Rusty-necklaced partridge has expanded to Lixian, Beidao and Haiyuan, which result in peripheral populations, paralleling with *A. chukar* along the Liupan Mountains (Fig. 1). Introgressive hybridization between the two species was detected in the contact zone (CHEN *et al.* 1999, LIU *et al.* 2006). These populations provided the opportunity to investigate the genetic diversity of peripheral populations, compared to that of central populations.

Mitochondrial DNA (mtDNA), particularly focusing on fast-evolving segments of the noncoding control region, has been extensively employed to assess evolutionary questions (STANLEY *et al.* 1996, BONATTO & SALZANO 1997, VILA *et al.* 1997). Recently, the development of hypervariable genomic markers, microsatellites (GOLDSTEIN & SCHLOTTERER 1999) allowed the inferring of additional details on evolutionary processes and population structure (BALDING *et al* 2001). Here we examine the difference of genetic diversity in relation to geographic position (peripheral or central), using both mtDNA control-region sequences and nuclear microsatellites. To eliminate the effects of genetic variation between subspecies, we only analyze one subspecies, *A. magna lanzhouensis*. There were two aims: (1) assess whether maternal mtDNA and biparental microsatellite markers described concordant population genetic diversity; and (2) compare the genetic diversity between peripheral and central populations.

188

MATERIALS AND METHODS

Sample collection and laboratory methods

A total of 82 samples of eight populations in rusty-necklaced partridge are collected from the following localities: Lanzhou, Dingxi, Jingyuan, Haiyuan, Huining, Beidao, Lixian and Wushan (Fig. 1). Wild samples were collected during consecutive hunting seasons. Liver samples were dissected from birds and stored in 95% ethanol immediately after removal. The methods of DNA extraction, PCR amplification and sequence of mtDNA control region genes referred to HUANG *et al* (2007*a*). The sequences were deposited in GenBank and the accession numbers are from DQ157593 to DQ157619. These are just from HUANG *et al* (2007*a*).

All samples were genotyped by PCR amplifications of eight microsatellites: MCW135 (5'-ATA TGC TGC AGA GGG CAG TA-3', 5'-CAT GTT CTG CAT TAT TGC TCC-3', annealing temperature = 45 °C), MCW207 (5'-GAT CCT TAC AGC CTG CAA TGC-3', 5'-ATA CTG TTG GAA GAT GTA TGC G-3', 60 °C), MCW295 (5'-ATC ACT ACA GAA CAC CCC TCT C-3', 5'-TAT GTA TGC ACG CAG ATA TC-3', 50 °C), MCW323 (5'-GAA ATG GTA CAG TGC AGT TGG-3', 5'-TGA ATT CTC TCG GCT TCC ATC-3', 60 °C), that were isolated originally from the chicken (*Gallus gallus*), and AB121114 (5'-GAC TAG TAG TGA AGA CTG TT-3', 5'-AGA TTT CTG GCT TCT GCA-3', 52 °C), AB063167 (5'-GTC ACA CAC TGT ATC ATA CT-3', 5'-GTG ATC TCA GTG TTT ATC TT-3', 55 °C), AB035840 (5'-TGC ACC AAT CCC AGC TGT TT-3', 5'-ACA ATG GAA AGT GGG GTT C-3', 55 °C), AB063153 (5'-CAT AAC TGG GAT ATT GTT TA-3', 5'-ACA ACC ACT TCT CCA GCT A-3', 52 °C) that from common quail (*Coturnix coturnix*), which were obtained from GenBank. The PCR products were denatured at 94 °C 5 min using Dextran blue formamide solution. After polyacrylamide gel electrophoresis, the migration rate fragment size was determined using Bandscan 4.30 software (http://moleco.sjtu.edu.cn), with the marker pUC19 DNA/Msp I (Hpa II).



Fig. 1. Rusty-necklaced partridge sampling sites: 1 = Lanzhou, 2 = Jingyuan, 3 = Haiyuan, 4 = Dingxi, 5 = Huining, 6 = Wushan, 7 = Beidao, 8 = Lixian

Sequence analysis

All sequences were aligned using Clustal X (THOMPSON *et al.* 1997). Arlequin2.0 (SCHNEIDER *et al.* 2002) was used to compute the number of haplotypes in populations, number of polymorphic sites. DnaSP4.0 (ROZAS *et al.* 2003) was used to calculate population haplotype diversity (*H*), nucleotide diversity (π) and mean number of pairwise differences (*K*). Arlequin2.0 (SCHNEIDER *et al.* 2002) was used to compute pairwise population differentiation and to perform analysis of molecular variance (AMOVA, EXCOFFIER *et al.* 1992).

The software GENEPOP Version 3.2a (ftp://ftp.cefe.cnrs-mop-fr/pub/msdos/genepop) (RAY-MOND & ROUSSET 1995) was used to calculate allele frequencies, observed (H_0) and expected (H_E) heterozygosities. Deviations from Hardy–Weinberg equilibrium for each locus and each population were assessed using the Markov chain method, as implemented in GENEPOP 3.2a. Genetic differentiations were tested among all pairs of populations for all loci (GENEPOP 3.2a). F_{sT} values for population subdivision were also calculated using GENEPOP 3.2a according to WEIR and COCKERHAM (1984). Tests of genotypic differentiation, based on the G-based exact tests of GOUDET *et al* (1996), were also performed using this program.

RESULTS

Mitochondrial DNA haplotype and variability

A total of 458 nucleotides of the mtDNA control region were sequenced of all the samples. The mtDNA control-region sequence alignment showed 25 different haplotypes, defined by 27 polymorphic sites (Table 1). The number of observed haplotypes within populations ranged from three in Beidao to seven in Lanzhou (Table 2). The percentages of unique haplotypes were calculated by dividing the number of unique haplotypes by the total number of samples. Within each population, this percentage varied from 17.64% in Lanzhou to 37.50% in Wushan (Table 2). The most common haplotypes were M2 with 29 individuals from all the sampling sites (Table 1). Many allied haplotypes, however, were localized. Results of AMOVA showed that 12.25% of the total mtDNA genetic variability was distributed within, and 87.75% among populations ($\Phi_{ST} = 0.63$, P<0.01). Pairwise F_{ST} values test showed peripheral populations were significantly differentiated from central populations except Haiyuan and Huining (Table 3).

Nucleotide diversity among the eight populations varied from 0.0028 (Haiyuan) to 0.0069 (Dingxi, Table 2); and haplotype diversity ranged from 0.52 (Beidao) to 0.86 (Wushan, Table 2). The pairwise divergence between haplotypes (average k = 2.33) was lowest (k = 0.85) in partridges from Haiyuan population and highest (k = 3.18) in partridges from Dingxi population. Three peripheral populations (Lixian, Beidao and Haiyuan), possessed lower haplotype diversity (average 0.67) and nucleotide diversity (average 0.0030), compared to central geographic populations (average H = 0.80, $\pi = 0.0057$).

Table 1.	Sampling location <i>t al.</i> 2007 <i>a</i>). Haple	ns, numbers and frequency in the total population of the 25 mtL otype positions are aligned with the complete mtDNA D-loop	DNA haplotypes found in rusty-necklace partridge (HUANG sequence of <i>Alectoris</i> (RANDI & LUCCHINI 1998).
Haplo-	Number	Variable positions in sequences	Sampling location
type	(frequency, %)		(sample size)
		000112222222222222223333333333344	
		112091111222334444578901134899945	
		797300178348562456388040458347964	
M1	2 (2.4)	AAAACAGCAGCTTTCTTTTCTATTCTGATTTCCC	Lanzhou (1), Lixian (1)
M2	29 (35.4)		Huining (5), Beidao (1), Haiyuan (7), Lanzhou (6), Dingxi (2), Wushan (1), Lixian (3), Jingyuan (4)
M3	3 (3.6)	T.G	Lixian (3)
M4	1 (1.2)	AT	Lixian (1)
M5	5(6.1)	ATC	Beidao (5)
M6	1 (1.2)	TC	Beidao (1)
M7	2 (2.4)	A	Wushan (2)
M8	1 (1.2)	.GATA.	Wushan (1)
9M	1 (1.2)	C.T.CC.	Wushan (1)
M10	8 (9.7)	C	Lanzhou (3), Jingyuan (1), Haiyuan (1), Wushan (3)
M11	5(6.1)	CC	Dingxi (5)
M12	1 (1.2)	CC	Dingxi (1)
M13	2 (2.4)	GC	Dingxi (1), Huining (1)
M14	1 (1.2)	GT	Huining (1)
M15	1 (1.2)	GC	Huining (1)
M16	2 (2.4)	T ⁻	Huining (1), Haiyuan (1)
M17	1 (1.2)	C	Haiyuan (1)
M18	1 (1.2)	C.GG	Haiyuan (1)

		Table 1 (continued)	
Haplo-	Number	Variable positions in sequences	Sampling location
type	(frequency, %)		(sample size)
M19	2 (2.4)	DG	Haiyuan (2)
M20	3 (3.6)	GCCG	Jingyuan (3)
M21	2 (2.4)	CG.	Jingyuan (2)
M22	2 (2.4)		Lanzhou (2)
M23	3 (3.6)	ACT	Lanzhou (2), Dingxi (1)
M24	2 (2.4)	G.TC	Lanzhou (2)
M25	1 (1.2)	T	Lanzhou (1)

Microsatellites genetic diversity

HUANG, Z. H., LIU, N. F., CHEN, Y. K. & XIAO, Y. A.

The results of our PCR amplifications of the eight microsatellite loci in 82 rusty-necklaced partridge samples revealed a total of 54 alleles. The eight microsatellites were polymorphic in the partridge samples, with the exception of locus MCW207, which was monomorphic in the all samples. Allele frequencies at microsatellites were calculated for all individuals. Values of observed heterozygosity (H_0) ranged from 0.20 (Lixian) to 0.75 (Jingyuan), and values of expected heterozygosity (H_E) varied from 0.31 (Lixian) to 0.59 (Wushan) (Table 2). The averages of the eight geographic populations of rusty-necklaced partridge are 0.45 for H_o and H_e. Significant allele frequency differences were detected among all pairwise comparisons for the eight ps over all loci (P <0.001). Probability tests for departure from Hardy-Weinberg performed in each population and cross each locus show that five loci (AB063153, MCW295, CW323, AB121114, AB035840) in each population were in equilibrium P > 0.05), and the MCW135 locus in the Lanzhou and Beidao populations was not equilibrium (P < 0.05). The multilocus test performed for Beidao population showed a heterozygote deficit, but the difference was not significant (P > 0.05). Other populations showed heterozygote redundancy, which was significant in populations Haiyuan and Jingyuan (P<0.05). Microsatellite genetic diversity was also significantly partitioned among the eight population (average multilocus $F_{ST} = 0.309$, P < 0.01). Pairwise F_{ST} values were significant between peripheral and central populations except Lixian and Wushan (Table 3).

Compared to central geographic populations, Lixian, Beidao and Haiyuan exhibited

Acta zool. hung. 55, 2009

	Tuble 2. Huplotypes and generie diversity of the eight populations.							
Population	Sample size	Total haplotypes	Unique haplotypes	K*	π^*	H*	H _o	$H_{\rm E}$
Huining	9	5	2	2.17	0.0047	0.73	0.57	0.55
Wushan	8	5	3	3.14	0.0057	0.86	0.50	0.58
Jingyuan	10	4	2	2.47	0.0054	0.78	0.75	0.46
Lanzhou	17	7	3	2.60	0.0057	0.85	0.52	0.59
Dingxi	10	5	2	3.18	0.0069	0.76	0.48	0.41
Beidao	7	3	2	1.81	0.0039	0.52	0.22	0.32
Lixian	8	4	2	1.03	0.0023	0.78	0.34	0.38
Haiyuan	13	6	3	0.85	0.0028	0.72	0.20	0.31

Table 2. Haplotypes and genetic diversity of the eight populations.

*From HUANG et al 2007a.

low H_o (average 0.25) and H_E (average 0.34). A significant difference was found in H_o (t = 2.2443, p = 0.044) and in H_E (t = 4.15, P = 0.007) between peripheral and the central populations. Lixian population has the lowest observed heterozygosity (H_o = 0.20) and the lowest expected heterozygosity (H_E = 0.31), significantly different from heterozygosity values of all the other populations (P < 0.05; Wilcoxon's signed-rank test).

DISCUSSION

Though the mtDNA genome of animals is typically inherited in a uniparental (matrilineal) fashion and only has an effective population size one-fourth that of the nuclear genome (AVISE *et al.* 1987), the genetic diversities exhibited by

Table 3. Pairwise values of F_{ST} (microsatellites DNA, above the diagonal; mitochondrial DNA, be-
low the diagonal) among populations of rusty-necklaced partridges.

			011			···· F···	0	
Population	Huining	Beidao	Haiyuan	Jingyuan	Lanzhou	Dingxi	Wushan	Lixian
Huining		0.396*	0.264*	0.358*	0.281*	0.367*	0.198*	0.413**
Beidao	0.654**		0.194*	0.264*	0.287*	0.771**	0.180*	0.273*
Haiyuan	0.031	0.640**		0.147*	0.337*	0.284*	0.110*	0.373*
Jingyuan	0.150*	0.545**	0.082		0.392*	0.324*	0.103*	0.392*
Lanzhou	0.126*	0.504**	0.102*	0.041		0.343*	0.302*	0.485**
Dingxi	0.176*	0.629**	0.249*	0.081	0.138*		0.254*	0.464**
Wushan	0.245*	0.310*	0.192*	0.154*	0.080	0.299*		0.088
Lixian	0.294*	0.499**	0.285*	0.205*	0.146*	0.347*	0.127*	

*P < 0.05, **P < 0.01

mtDNA haplotypes and observed at microsatellites loci were accordant. The three peripheral populations, Lixian, Beidao and Haiyuan possessed lower nucleotide diversity (average $\pi = 0.0030$), haplotype diversity (average H = 0.67), and values of observed heterozygosity (average H₀ = 0.25) and expected heterozygosity (average H_E = 0.34), while central populations owned higher genetic diversity (average $\pi = 0.0057$, H = 0.80, H_E = 0.52, H₀ = 0.58).

Many authors believe that the peripheral populations often have reduced levels of genetic variability relative to central populations (LESICA & ALLERDORF 1995, GARCIA-RAMOS & KIRKPATRICK 1997, WANG et al. 2001). Our results support this hypothesis, since compared to central populations, the three peripheral populations exhibited lower genetic diversity. Populations located at range margins are more isolated from sources of immigrants and are thus more prone to genetic bottlenecks (KARRON 1987, ROWE & BEEBEE 2003), a situation that should deplete neutral genetic variation (GARNER et al. 2004). The genetic diversity of a population is related to the degree of isolation. Low levels of genetic diversity can be expected in populations at range limits as a result of low levels of immigration and high levels of genetic drift (e.g. SOULÉ 1973, HOFFMANN & BLOWS 1994). Rusty-necklaced partridge is a species indicative of arid and semiarid environments in northwestern China, while forest and farmland are generally avoided. This could explain the lower genetic diversity of the Haiyuan population, because it is isolated from other populations by farmlands, preventing gene flow. HUANG et al. (2007b) observed that the population genetic diversity of rusty-necklaced partridge was negatively correlated with the rainfall. Based on this environmental factor, natural selection could lead to a lower genetic diversity. Indeed, the Lixian and Beidao populations belong to wet areas with average annual rainfall of 510.0±126.2 mm (n = 40) and 547.8 \pm 130.5 mm (n = 40), a habitat little favorable for rustynecklaced partridges, and possessed the lowest genetic diversities (Table 2).

Acknowledgements – We would like to thank Dr. ETTORE RANDI for providing primers and technical assistance, and Dr. PENG HOU, Mr. MING WEI and Dr. GUOJU XIAO for assistance in obtaining samples. We are especially grateful to the two anonymous reviewers for helpful comments on a previous version of this manuscript. This work was supported by National Natural Science Foundation of China (Nos 30530130, 30760036).

REFERENCES

AVISE, J. C., ARNOLD, J., BALL, R. M., BERMINGHAM, E., LAMB, T., NIEGEL, J. E., REEB, C. A. & SAUNDERS, N. C. (1987) Intraspecific phylogeography: the mitochondrial DNA bridge be-

tween population genetics and systematics. *Annual Review of Ecology and Systematics* **18**: 489–522.

- BALDING, D. J., BISHOP, M. & CANNINGS, C. (2001) Handbook of Statistical Genetics. John Wiley & Sons, Chichester, UK, pp. 980–1013.
- BARRETT, S. C. H. & KOHN, J. R. (1991) Genetic and evolutionary consequences of small population size in plants: implications for conservation. Pp. 3–30. *In*: FALK, D. A. & HOLSINGER, K. E. (eds): *Genetics and Conservation of Rare Plants*. Oxford Univ. Press.
- BONATTO, S. L. & SALZANO, F. M. (1997) A single and early migration for control-region sequences in a marine mammal: Implications for conservation and biogeography of Steller sea lions (Eumetopias jubatus). *Journal of Mammalogy* 77: 95–108.
- CASSEL, A. & TAMMARU, T. (2003) Allozyme variability in central, peripheral and isolated populations of the scarce heath (Coenonympha hero: Lepidoptera, Nymphalidae); implications for conservation. *Conservation Genetics* 4: 83–93.
- CHEN, Q., CHANG, C., LIU, N. F., RANDI, E. & LUCCHINI, V. (1999) Mitochondrial DNA introgression between two parapatric species of Alectoris. Acta Zoologica Sinica 45: 456–463.
- ECKERT, C. G., SAMIS, K. E. & LOUGHEED, S. C. (2008) Genetic variation across species' geographical ranges: the central-marginal hypothesis and beyond. *Molecular Ecology* 17: 1170–1188.
- EXCOFFIER, L., SMOUSE, P. E. & QUATTRO, J. M. (1992) Analysis of molecular variance inferred from metric distances among DNA haplotypes: application to human mitochondrial DNA restriction data. *Genetics* 131: 479–491.
- GARCIA-RAMOS, G. & KIRKPATRICK, M. (1997) Genetic models of adaptation and gene flow in peripheral populations. *Evolution* **51**: 21–28.
- GARNER, T. W. J., PEARMAN, P. B. & ANGELONE, S. (2004) Genetic diversity across vertebrate species' range: a test of central–peripheral hypothesis. *Molecular Ecology* 13: 1047–1053.
- GOLDSTEIN, D. B. & SCHLOTTERER, C. (1999) *Microsatellites Evolution and Applications*. Oxford University Press, New York, pp. 49–65.
- GOUDET, J., RAYMOND, M., DE MEEU, S. T. & ROUSSET, F. (1996) Testing differentiation in diploid populations. *Genetics* 144: 1933–1940.
- HARTL, D. L. & CARK, A. G. (1997) Principles of Population Genetics. 3rd edition. Sinauer Associate, Inc. Sunderland, Massachusetts, pp. 267–312.
- HOFFMANN, A. A. & BLOWS, M. W. (1994) Species borders: ecological and evolutionary perspectives. Trends of Ecology and Evolution 9: 223–227.
- HOU, P., WEI, M., ZHANG, L. X. & LIU, N. F. (2002) Genetic structure of edge population in Przewalski's rock partridge Alectoris magna. *Acta Zoologica Sinica* 48: 333–338.
- HUANG, Z. H., LIU, N. F., LUO, S. X. & LONG, J. (2007a) Phylogeography of rusty-necklaced partridge (Alectoris magna) in northwestern China. *Molecular Phylogenetics and Evolution* 43: 379–385.
- HUANG, Z. H., LIU, N. F., LUO, S. X., LONG, J. & XIAO, Y. A. (2007b) Ecological genetics of rusty-necklaced partridge (Alectoris magna): environmental factors and population genetic variability correlations. *Korean Journal of Genetics* 29: 115–120.
- HUANG, Z. H., LIU, N. F. & ZHOU, T. L. (2005) A comparative study of genetic diversity of peripheral and central populations of chukar partridge from northwestern China. *Biochemical Genetics* **43**: 613–621.
- JAIN, S. K., RAI, K. N. & SINGH, R. S. (1981) Population biology of Avena XI. Variation in peripheral isolates of A. barbata. *Genetian* 56: 233–215.
- KARRON, J. D. (1987) A comparison of levels of genetic polymorphism and self-incompatibility in geographically restricted and widespread plant congeners. *Evolutionary Ecology* 1: 47–58.

- KIRKPATRICK, M. & BARTON, N. H. (1997) Evolution of a species range. *The American Naturalist* **150**: 1–23.
- LAMMI, A., SIIKAMAMI, P. & MUSTAJARVI, K. (1999) Genetic diversity, population size, and fitness in central and peripheral populations of a rare plant Lychnis viscara. *Conservation Biology* **13**: 1069–1078.
- LESICA, P. & ALLERDORF, F. W. (1995) When are peripheral populations valuable for conservation? Conservation Biology 9: 753–760.
- LIU, N. F., HUANG, Z. H. & WEN, L. Y. (2004) Subspecies divergence of Przewalski's rock partridge (Alectoris magna) description of a new subspecies. *Acta Zootaxonomica Sinica* **29**: 600–605.
- LIU, N. F., WEN, L. Y., HUANG, Z. H. & HOU, P. (2006) Introgressive hybridization between Alectoris magna and A. chukar in the Liupan Mountain Region. Acta Zoologica Sinica 52: 153–159.
- LIU, N. F. (1992) Ecology of Przewalski's Rock Partridge (Alectoris magna). *Gibier Faune Sauvage* 9: 605–615.
- NEI, M., MARUYAMA, T. & CHAKRABORTY, R. (1975) The bottleneck effect and genetic variability in populations. *Evolution* 29: 1–10.
- PETITET, R. J., MOUSADIK, A. E. & POAS, O. (1998) Identifying populations for conservation on the basis of genetic markers. *Conservation Biology* 12: 844–855.
- RANDI, E (1996) A mitochondrial cytochrome *b* phylogeny of the Alectoris partridges. *Molecular Phylogenetic Evolution* **6**: 214–27.
- RANDI, E. & LUCCHINI, V. (1998) Organization and evolution of the mitochondrial DNA control region in the avian genus Alectoris. *Journal of Molecular Evolution* 47: 449–462.
- RANDI, E., TABARRONI, C., RIMONDI, S., LUCCHINI, V. & SFOUGARIS, A. (2003) Phylogeography of the rock partridge (Alectoris graeca). *Molecular Ecology* **12**: 2201–2214.

RAYMOND, M. & ROUSSET, F. (1995) Genepop version 3.2: population genetics software for exact tests and ecumenicism. *Journal of Heredity* 86: 248–249.

- ROWE, G. & BEEBEE, T. J. C. (2003) Population on the verge of a mutational meltdown? Fitness costs of genetic load for an amphibian in the wild. *Evolution* **57**: 177–181.
- ROZAS, J., SÁNCHEZ-DELBARRIO, J. C., MESSEGUER, X. & ROZAS, R. (2003) DNASP, DNA polymorphism analyses by the coalescent and other methods. *Bioinformatics* 19: 2496–2497.
- SAFRIEL, U. N., VOLIS, S. & KARK, S. (1994) Core and peripheral populations and global climate changes. *Israel Journal of Botany Sciences* 42: 331–345.
- SCHNEIDER, S., ROESSLI, D. & EXCOFFIER, L. (2002) Arlequin Version 2.0. A Software for Population Genetics Data Analysis Genetics and Biometry Laboratory, Department of Anthropology and Ecology, University of Geneva, Geneva, Switzerland.
- SOULÉ, M. (1973) The epistasis cycle: a theory of marginal populations. *Annual Review of Ecology* and Systemtatics 4: 165–187.
- STANLEY, H. F., CASEY, S., CARNAHAM, J. M., GOODMAN, S., HARWOOD, J. & WAYNE, R. K. (1996) Worldwide patterns of mitochondrial DNA differentiation in the harbor seal (Phoca vitulina). *Molecular Biology and Evolution* 13: 368–382.
- THOMPSON, J. D., GIBSON, T. J., PLEWNIAK, F., JEANMOUGIN, F. & HIGGINS, D. G. (1997) The Clustal X windows interface flexible strategies for multiple sequence alignment aided by quality analysis tool. *Nucleic Acid Research* 24: 4876–4882.
- TIGERSTEDT, P. M. A. (1973) Studies on isozyme variation in marginal and central populations of Picea abies. *Hereditas* 75: 47–60.
- VILA, C., SAVOLAINEN, P., MALDONADO, J. E., AMORIM, I. R., RICE, J. E., HONEYCUTT, R. L., CRANDALL, K. A., LUNDEBERG, G. & WAYNE, R. K. (1997) Multiple and ancient origins of the domestic dog. *Science* 276: 1687–1689.

- WANG, Y. Q., ZHANG, Z. B. & XU, L. X. (2001) The genetic diversity of central and peripheral populations of ratlike hamster (Cricetulus triton). *Chinese Science Bulletin* **46**: 1644–1650.
- WEIR, B. S. & COCKERHAM, C. C. (1984) Estimating F-statistics for the analysis of population structure. *Evolution* 38: 1358–1370.
- WENDEL, P. F. & PARKS, C. R. (1985) Genetic diversity and population structure in Camellia japonica L. (Theaceae). American Journal of Botany 72: 52–65.
- YEH, F. C. & LAYTON, C. (1979) The organization of genetic variability in central and marginal population of lodgepole pine Pinus contorta spp. latifola. *Canadian Journal of Genetics and Cytol*ogy 21: 87–503.

Revised version received January 8, 2009, accepted April 14, 2009, published May 29, 2009